

19546 pIMAGO®-biotin Phosphoprotein Detection Kit, for Microplate Blot, Fluor 680-based detection (400 wells)

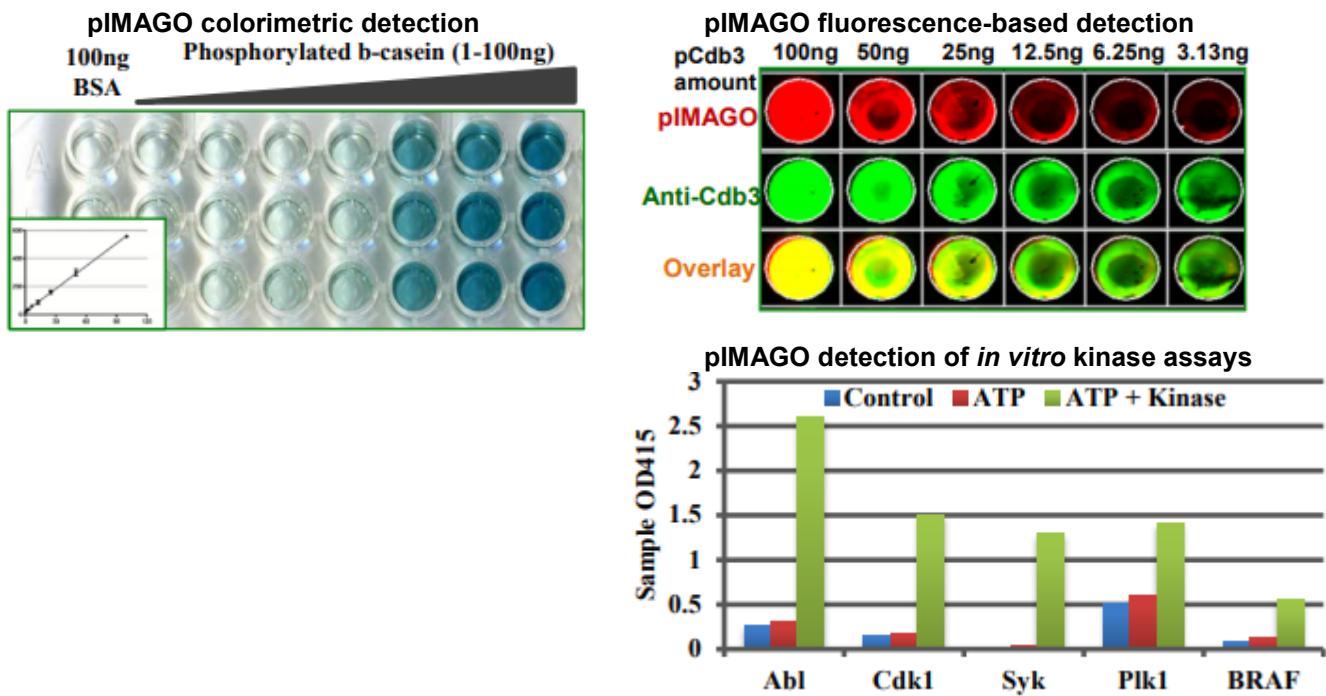
Storage Temperature 2-8°C

Introduction

pIMAGO is a universal phosphoprotein detection technology that enables sensitive and specific recognition of phosphorylated molecules. Unlike phospho-antibodies, the binding is not biased by amino acid sequence, and therefore can be used for detection of any phosphorylation event on any protein site. pIMAGO detection protocol resembles a simple ELISA procedure and can be easily incorporated by any laboratory. Due to its small size, pIMAGO® can be multiplexed with antibodies for simultaneous detection of phosphorylation and total protein amount.

Results

pIMAGO-based detection and quantitation of phosphorylated β -casein signal compared to non-phosphoprotein BSA (*left figure*). Multiplexed detection of phospho-Cdb3 using pIMAGO in the 700 channel and anti-Cdb3 antibody in the 800 channel (*right figure*). pIMAGO-based kinase assays of 5 kinases and their substrates, including control, ATP, and ATP + Kinase wells (*bottom figure*).



Kit components

- 400 μ L of pIMAGO reagent
- 400 μ L of avidin-Fluor 680
- 40 ml of Binding buffer
- 240 mL of Blocking buffer
- 240 mL of TBST (Tris-buffered saline with 0.1% Tween 20)
- 240 mL of pIMAGO buffer
- 200 μ L of control phosphoprotein (β -casein; mix 5 μ L with 95 μ L Binding buffer)
- 10 flat-bottom clear 96-well plates



Protocol

1. Binding of samples to the 96-well plate

Prepare a protein solution of your sample (phosphoprotein or substrate of interest) in **Binding buffer**. A dephosphorylated (e.g. alkaline phosphatase-treated or without ATP) form of the sample should also be prepared as a reference. If protein amount is known, use 10 to 500 ng of the protein or mixture of proteins per 100 µL of **Binding buffer** per well. Add 100 µL of the mix into each well on a 96-well plate. Incubate overnight at 4°C at 400-600 rpm to bind the proteins to the plate. As a positive control, in a separate well mix 5 µL of the provided phosphoprotein with 95 µL of Binding buffer.

2. Blocking the wells

Remove solution from wells, add 150 µL of the **Blocking buffer** into each well and incubate 2-3 minutes. Remove the solution and add 150 µL of the **Blocking buffer** again and incubate for 30 min while shaking at 400-600 rpm at room temperature.

At this stage, any additional manipulations can be carried out (e.g. kinase/phosphatase assay, inhibitor screening, etc.). Make sure to wash the wells 3x with the 1x TBST after each manipulation.

3. pIMAGO incubation

In a clean tube, prepare a 1 to 100 mixture of the **pIMAGO reagent** in the **pIMAGO buffer** (1 µL of the reagent for every 100 µL of buffer). Empty the wells and add 100 µL per well of the prepared pIMAGO mix. Incubate 1 hour at 400-600 rpm at room temperature.

4. Washing the wells

Empty the wells and add 150 µL of the **pIMAGO buffer** into each well; incubate 2-3 minutes at 400-600 rpm. Remove the buffer and repeat the washing step two more times with the **pIMAGO buffer** for a total of 3 washes. Remove the solution and incubate the wells with 150 µL of the **Blocking buffer** for 15 min at 400-600 rpm at room temperature.

5. Incubation with avidin-Fluor

In a clean tube, prepare 1 to 100 mixture of **avidin-Fluor** in the **Blocking buffer** (1 µL of **avidin-Fluor** in 100 µL of **Blocking buffer**). Empty the wells and add 100 µL per well of the prepared avidin-HRP in blocking solution. Incubate the plate for 1 hour at 400-600 rpm at room temperature.

6. Washing the wells

Empty the wells and add 150 µL of **1x TBST** into each well; incubate 2-3 minutes at 400-600 rpm and remove the solution. Repeat the washing step with TBST two more times for a total of three washes. Empty the wells.

7. Signal detection

The signal can be detected using any fluorescence imager with an appropriate filter. The number listed with the provided avidin-Fluor reagent represents the absorption maximum (emission wavelength) for that fluorophore.

References

“Phosphorylation assay based on functionalized soluble nanopolymer”. Iliuk, A.; Martinez, J.; Hall, M. C.; Tao, W. A., *Anal. Chem.* **2011**, 83, 2767-2774. PMID: 21395237

Legal Information

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Precautions and Disclaimer

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