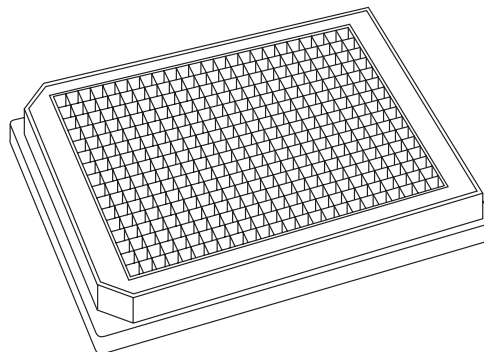


## User Guide

# MultiScreen®<sub>HTS</sub> 384-Well Filter Plate

For research use only. Single use only.



## Indications for Use

For use in automated high throughput applications including sample preparation and radiometric filter-binding assays.

## Introduction

MultiScreen®<sub>HTS</sub> 384-well plates are non-sterile, single use filter plates that can be used to perform high throughput biological assays or to prepare samples for subsequent analysis. The filter plate can be used in a variety of biological assays including receptor-ligand binding (RLB) assays, assays that measure enzyme activity (e.g., kinases, phosphatases, proteases, etc.), and general sample prep.

The MultiScreen®<sub>HTS</sub> 384-well plate is designed to be compatible with robotic workstations for automated manipulation and can be used with either vacuum or centrifugal filtration modes. These filter plates are compatible with 384-well receiver plates for the collection and analysis of filtered samples.

## Materials Required

The user must supply the following materials in order to use the MultiScreen®<sub>HTS</sub> 384-well filter plate.

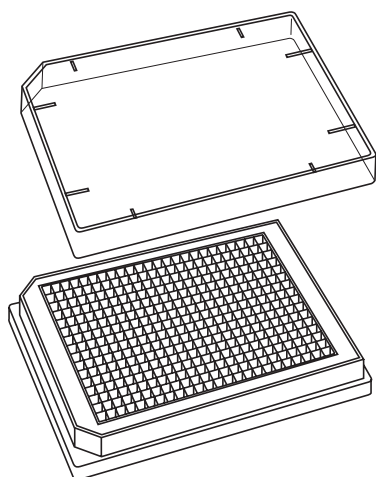
### Reagents

- Application appropriate aqueous buffers, biological components, and other assay materials.

### Equipment

- Manual pipettors or robotic liquid handling systems for transferring of liquid into filter plates.
- Vacuum manifold (Cat. No. MSVMHTS00) or centrifuge with a swinging bucket rotor and multiwell plate carrier.
- Solid bottom 384-well assay plates (if collecting filtrate).

## Diagram of MultiScreen®<sub>HTS</sub> 384-well Filter Plate with Lid



## Chemical Compatibility

R= Resistant, N= Non-Resistant,  
T= User should test in application

Chemical	Compatibility	
	Membrane	Glass
<b>Acids</b>		
Acetic Acid (Glacial)	N	R
Acetic Acid (25%)	R	R
Sulfuric Acid (90%)	N	N
<b>Scintillants</b>		
MicroScint-O™ Scintillant	R	R
MicroScint® 40 Scintillant	R	R
Opti-Fluor® Scintillant	R	R
OptiPhase™ Supermix™ Scintillant	N	R
Ultima Gold™ Scintillant	R	R
<b>Organic Solvents</b>		
Acetone	N	N
Acetonitrile	N	N
Tetrahydrofuran	N	N
Diethyl Ether	N	N
DMSO	N	N
Glycerol	R	R
Methanol	T	T
<b>Aqueous Solvents</b>		
Sodium Dodecyl Sulfate (SDS)	R	R
Tween® Surfactant	R	T
Phosphate Buffered Saline (PBS)	R	R
Phosphoric Acid (1%)	R	R
Hydrochloric Acid, HCl (1 N)	R	R
Sodium Hydroxide, NaOH (1 N)	R	R
Acetonitrile (35%)	N	N
DMSO (5%)	R	R

## Usage Guidelines

- The MultiScreen®<sub>HTS</sub> 384-well filter plate can be operated either in a vacuum or centrifugal pressure mode at temperatures between 4 °C and 37 °C. Recommended vacuum pressure is 8–15 inHg for glass fiber plates and membrane plates. Recommended centrifugal force is 500–1,000 × g.
- In either vacuum or centrifugal mode, samples can be collected in a solid bottom receiver (assay) plate. In vacuum mode, sample or (wash) buffer(s) can be collected into a solid bottom receiver plate or transferred directly to waste.
- Although the MultiScreen®<sub>HTS</sub> 384-well plate has been validated for use with extended incubations of aqueous media and scintillants, care should be taken to protect working surfaces from contamination in case of leakage.
- When doing radiometric inplate analysis with the Wallac MicroBeta® TriLux® liquid scintillation counter, plate carriers should be taped.
- Extended incubations with solutions containing high levels of surfactant may lead to drip out; test the plate before use in experiments.
- The MultiScreen®<sub>HTS</sub> 384-well filter plate with glass fiber is compatible with polyethyleneimine (0.5% PEI) pre-treatment. **Do not allow PEI to dry!** For those applications requiring this step, wash twice with binding buffer to complete preparation of plates after initial pretreatment for 2–16 hours at 4 °C.
- Dry glass fiber plates after filtration if adding scintillant. After adding scintillant, wait at least 1 hour before reading to allow scintillant to fully coat the filter.
- Cover the plate with the lid while incubating or use some other means to reduce evaporation, especially if incubating for more than 1 hour or at elevated temperatures.

## Limitations

Operating Temperature Range	4 °C - 37 °C
Maximum Drying Temperature	50 °C
Maximum Vacuum Pressure	20 inHg
Maximum Centrifugal Force	1,000 × g

## Procedure

The following sections outline the use of the MultiScreen<sup>®</sup><sub>HTS</sub> 384-well filter plate.

### Sample Addition

Pipette the appropriate amount of test sample (1 to 100 µL) into each well of the filtration plate. For best results when adding multiple solutions to the well, add the solution with the largest volume first, and end by adding the solution containing the smallest volume.

### In-Plate Incubation

The MultiScreen<sup>®</sup><sub>HTS</sub> 384-well filter plate is designed to permit in-plate incubations of various liquids, thereby eliminating the need to perform assays in a separate plate prior to transferring the sample to the filter plate. Due to the low volumes typically incubated in the MultiScreen<sup>®</sup><sub>HTS</sub> 384-well filter plate (20–100 µL), it is recommended that the plate lid be used during extended incubations or incubations that are carried out at elevated temperatures (e.g., at or above 37 °C).

### Sample Filtration

Samples can be filtered using either centrifugal or vacuum modes. In centrifugal mode, a 384-well receiver plate is positioned under the filter plate to collect the filtrate. Vacuum filtration should be performed using the MultiScreen<sup>®</sup><sub>HTS</sub> Vacuum Manifold (Cat. No. MSVMHTS00). In vacuum mode, the filtrate is either collected in a receiver plate or it is transferred directly to waste.

### Filtrate Collection

The filter plate is designed to be used in conjunction with a SLAS (Society of Laboratory and Automation Screening, formally SBS/ANSI) designed solid bottom 384-well receiver (assay) plate. When stacked on top of a 384-well receiver plate, the MultiScreen<sup>®</sup><sub>HTS</sub> 384 well plate will direct the contents of each well into the corresponding well in the receiver plate.

For centrifugation, stack the filter plate on top of a receiver plate and then place the assembly into the plate carrier of any standard centrifuge with swinging bucket rotors.

In vacuum mode, stack the filter plate on top of the receiver plate and place the assembly in the base of the vacuum manifold. The collar fits over the assembly. See the MultiScreen<sup>®</sup><sub>HTS</sub> Vacuum Manifold User Guide for details.

**NOTE:** Optimal vacuum levels can vary among assays. Typically, the lowest vacuum setting that gives the desired performance should be used.

## Washing of Samples to Waste

For optimal performance during multiple washes of retentate sample (i.e., flow to waste), vacuum pressures of 12–15 inHg are recommended, depending on filter type.

## Automated Operation Guidelines

The MultiScreen<sup>®</sup><sub>HTS</sub> 384-well filter plate is designed to be used with automated liquid handling systems and robotic workstations that are compatible with SLAS standard 384-well plates. Integration of vacuum filtering capability may require some customization or be impossible on some systems.

## Radiometric Assay Counting Guidelines

Follow the guidelines below for inplate measurement of radioactivity.

### Whole Plate Scintillation Counting

Radioactivity (scintillation counting) can be measured directly in the MultiScreen<sup>®</sup><sub>HTS</sub> 384-well filter plate. The design allows the plate to be analyzed from the top (top counting), from the bottom (bottom counting), or from both sides simultaneously (coincidence counting). For best signal to noise ratios using the Wallac MicroBeta<sup>®</sup> TriLux<sup>®</sup> liquid scintillation counter, count in the coincidence mode. Scintillation counting should be performed after at least one hour of exposure to the liquid scintillant cocktail. In some instances, longer incubations (up to 24 hours) may result in even higher counting efficiencies. Because different assays have different counting criteria and levels of signal, optimum counting mode and time should be determined by the operator.

### Radioisotope Guidelines

Low energy isotopes such as <sup>14</sup>C, <sup>3</sup>H, <sup>125</sup>I, <sup>33</sup>P, and <sup>35</sup>S require the use of 5–10 µL of scintillation cocktail per well. For optimal results and consistency, the scintillation cocktail should be allowed to mix with the sample for at least an hour before counting. This time is required to allow for complete mixing of the radiolabeled materials into the scintillation cocktail.

## Specifications

### Materials of Construction

Base Plate	Styrene Acrylonitrile (SAN) top: White; bottom: clear
Plate Lid	Polystyrene
Support	Polyester (glass fiber only)
Membrane	PVDF

See "Product Ordering" section for filter types.

### Dimensions

Plate assembly length	127.8 mm (5.03 in.)
Plate assembly width	85.5 mm (3.366 in.)
Plate assembly depth (without lid)	14.4 mm (0.567 in.)
Plate assembly depth (with lid)	16.9 mm (0.665 in.)
Approximate filter surface area	0.09 cm <sup>2</sup> (0.014 in <sup>2</sup> )
Well Depth	approx. 10.9 mm (0.429 in.)

### Filtration parameters

Recommended vacuum pressure	8-15 inHg for glass fiber and membrane plates
Recommended centrifugal force	500-1,000 x g
Recommended sample volume per well	20-100 µL

## Product Ordering

Purchase products online at [SigmaAldrich.com](https://SigmaAldrich.com).

Membrane/Filter Type	Qty	Cat. No.
Hydrophilic 0.45 µm Durapore® PVDF	10/pk	MZHVN0W10
Hydrophilic 0.45 µm Durapore® PVDF	50/pk	MZHVN0W50
Glass Fiber B	10/pk	MZFBN0W10
Glass Fiber B	50/pk	MZFBN0W50
Glass Fiber C	10/pk	MZFCN0W10
Glass Fiber C	50/pk	MZFCN0W50

### Accessories

	Cat. No.
MultiScreen® <sub>HTS</sub> Vacuum Manifold	MSVMHTS00
Chemical Duty Pump	
115 Volts, 60 Hz	WP6111560
220 Volts, 50 Hz	WP6122050
Plate Sealing Tape, 100 Sheets	
Clear	MATAHCL00
White	MATAH0P00
Vacuum Flask, 1 L	XX1014705

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