Microscopy

New fuchsin (C.I. 42520)

for microscopy Certistain®



In Vitro Diagnostic Medical Device



This staining dye "New fuchsin (C.I. 42520) - for microscopy Certistain® is used for human-medical cell diagnosis and serves the purpose of the bacteriological and histological investigation of sample material of human origin. It is a dry staining dye that is used to prepare a staining solution, that when used together with other in vitro diagnostic products from our portfolio makes target structures (by fixing, where necessary embedding, staining with the above new fuchsin solution, counterstaining, mounting) in bacteriological and histological specimen materials, for example smears of enriched bacterial cultures or histological sections of e.g. the lung, evaluable for diagnostic purposes.

Principle

New fuchsin belongs to the group of triaminotriphenylmethane dyes.

The dye is used in Ziehl-Neelsen and Gram staining as well as for methods with Schiff's reagent.

The cell wall of mycobacteria has a high proportion of wax and lipids and hence absorbs dyes only very slowly. As a measure to enhance the absorption of the fuchsin dye and thus the formation of the mycolate-fuchsin complex in the cell wall, the carbolfuchsin solution applied to the specimen is normally heated to evaporation point. If the Kinyoun carbolfuchsin solution is used, the heating of the solution is dispensable. Consequently, the release of hazardous phenolic vapors will be avoided.

Once the mycobacteria have absorbed the fuchsin dye, it is virtually impossible to decolorize them again, even when they are intensively treated with a decolorizing solution such as e.g. hydrochloric acid in ethanol. Accordingly, mycobacteria are termed as acid- and alcohol-fast for staining, and are stained red in the microscopic visualization. Correspondingly, all non-acid-fast microorganisms are counterstained with an appropriate dye. In the present operating instructions is counterstained accordance with methylene blue.

Pretreatment of the specimens with Sputofluol® dissolves the bacteria from the surrounding viscid sputum and cell material. Sputofluol® also has a disinfectant effect, with the result that any microorganisms that are present are killed off.

Sample material

Smears of bacteriological material that have been air-dried, heat-fixed, and pretreated with Sputofluol® like sputum, smears from fine needle aspiration biopsies (FNAB), rinses, imprints, effusions, pus, exsudates, liquid and solid cultures

Sections of formalin-fixed tissue of human origin, embedded in paraffin (3 - 4 μ m thick paraffin sections)

Reagents

Cat. No. 1.05226.0100 New fuchsin (C.I. 42520) for microscopy Certistain®

100 g

Color Index No.: 42520
Color Index Name: Basic violet 2

Also required:

Cat. No.	100206	Phenol GR for analysis ACS,Reag. Ph Eur	250 g, 1 kg
Cat. No.	100327	Hydrochloric acid in ethanol for microscopy	1 I, 5 I
Cat. No.	100971	Ethanol 96% suitable for use as excipient EMPROVE® exp Ph Eur,BP	1 l, 2.5 l, 10 l
Cat. No.	101287	Löffler's methylene blue solution for microscopy	100 ml, 500 ml, 2.5 l

Alternatively:

Instead of the combination of single reagents, the staining kit 1.00497.0001 can be used:

Cat. No. 1.00497.0001

Tb-color modified

1 set

Staining kit for the detection of mycobacteria (AFB) by hot staining method

Sample pretreatment

The sampling must be performed by qualified personnel.

Sputum

The mycobacteria should be pretreated with Sputofluol® to dissolve them from mucus and cellular structures. In this process, the active ingredient hypochlorite dissolves the organic material by oxidation and gently releases the mycobacteria so that they can be processed further.

Reagent preparation: Preparation of Sputofluol® solution 15 % For preparation of approx. 100 ml solution mix:

Sputofluol®	15 ml
Distilled water	85 ml

Preparing sample material in centrifuge tubes:	
Sample	1 part (min. 2 ml)
Sputofluol® solution (15 % in distilled water)	3 parts
Shake vigorously	10 min
Centrifuge at 3000 - 4800 rpm	20 min
Decant supernatant Prepare smears of the sediment Air-dry	

Punctates, lavages, sediments

After appropriate enrichment measures, smear sample material on the slide and allow to air-dry.

Histological sections

Deparaffinize sections in the conventional manner and rehydrate in a descending alcohol series. Pretreatment with Sputofluol® is not necessary for specimens fixed with formalin.

Fixation

Specimens are fixed over a Bunsen burner flame (2 - 3 times, taking care to avoid excessive heating).

The specimens can also be fixed by heating at 100 - 110 °C in a drying cabinet or on a heating plate for 20 min.

Excessive temperatures or prolonged heating may involve a deterioration of the staining performance.

All samples must be treated using state-of-the-art technology. All samples must be clearly labeled.

Suitable instruments must be used for taking samples and their preparation. Follow the manufacturer's instructions for application / use.

Reagent preparation

Phenol solution, aqueous

For preparation of approx. 100 ml solution mix:

Phenol	5 g
Distilled water	100 ml
dissolve	

New fuchsin solution, alcoholic

For preparation of approx. 100 ml solution mix:

New fuchsin (C.I. 42520) Certistain®	3 g
Ethanol 96 %	100 ml
dissolve and filter	

New fuchsin working solution, phenolic

For preparation of approx. 100 ml solution mix:

New fuchsin solution, alcoholic	10 ml
Phenol solution, aqueous	90 ml
mix	

Diluted Löffler's methylene blue solution

For preparation of approx. 100 ml solution mix:

Löffler's methylene blue solution	10 ml
Distilled water	90 ml
mix	

The freshly prepared staining solutions should be filtered before use.

Procedure

Staining of mycobacteria on the staining rack

Deparaffinize histological slides in the conventional manner and rehydrate in a descending alcohol series.

Carefully heat-fix the smears

The stated times should be adhered to to guarantee an optimal staining result.

Slide with histological tissue (rinsed in distilled water) or Slide with fixed smear			
New fuchsin working solution, phenolic	cover completely, carefully heat 2 - 3 times from below with the Bunsen burner until steam forms Do not allow the staining solution to boil!	stain for 5 min in total	
Tap water	rinse	1 min	
Hydrochloric acid in ethanol	rinse until no further clouds of dye are produced	10 - 25 sec*	
Tap water	rinse immediately		
Diluted Löffler's methy- lene blue solution	counterstaining, cover completely and leave to react	1 min	
Tap water	rinse carefully		
Air-dry (e.g. over night or at 50 °C in the drying cabinet)			
Mount with Entellan® Neu or Neo-Mount® and cover glass			

depending on thickness of specimen

Covering with non-aqueous mounting media (e.g. Neo-Mount® or Entellan® Neu) and a cover glass is recommended for the storage of bacteriological specimens for several months. For this purpose, the stained specimens must be dried very well. When left unmounted, the stain remains stable for approx. 3 days, covered with immersion oil for just a few hours.

The use of immersion oil is recommended for the analysis of stained slides with a microscopic magnification >40x.

Result

Mycobacteria red Background light blue

Fvaluation

A positive result means "acid fast bacteria detected" and a negative result "acid fast bacteria not detected". It is not possible to tell whether the bacteria found are Mycobacterium tuberculosis or of a different species of mycobacterium. The vitality (active, inactive) of the bacteria can also not be determined.

If mycobacteria are detected, further analyses should be performed in specially equipped laboratories.

Trouble-shooting

A sufficient degree of heat-fixing using a Bunsen burner or in a heating cabinet is essential to prevent the infectious potential of the specimens and further proliferation of the bacteria.

Excessive temperatures or prolonged heating may involve a deterioration of the staining performance.

No staining of mycobacteria

The critical step of the mycobacteria-staining process is the decolorizing step, which can be influenced by the thickness of the specimen smear.

In addition, a freshly prepared solution of hydrochloric acid in ethanol is highly reactive, meaning that the result should be evaluated with caution. The incubation times stated in this protocol should be kept accurately in the decolorizing step, since otherwise false-negative results may ensue.

Technical notes

The microscope used should meet the requirements of a medical diagnostic laboratory.

When using automatic staining systems, please follow the instructions for use supplied by the supplier of the system and software.

The freshly prepared staining solutions should be filtered before use.

Remove surplus immersion oil before filing.

Diagnostics

Diagnoses are to be made only by authorized and trained personnel. Valid nomenclatures must be used.

Further tests must be selected and implemented according to recognized methods. Suitable controls (e.g. ISOSLIDE® AFB, Cat. No. 1.02560.0001) should be conducted with each application in order to avoid an incorrect result.

Storage

Store New fuchsin (C.I. 42520) - for microscopy Certistain® at +5 °C to +30 °C.

Shelf-life

New fuchsin (C.I. 42520) - for microscopy Certistain® can be used until the stated expiry date.

After first opening of the bottle, the contents can be used up to the stated expiry date when stored at +5 °C to +30 °C.

The bottles must be kept tightly closed at all times.

Additional instructions

For professional use only.

In order to avoid errors, the application must be carried out by qualified personnel onlv.

National guidelines for work safety and quality assurance must be followed.

Microscopes equipped according to the standard must be used.

If necessary use a standard centrifuge suitable for medical diagnostic laboratory.

Protection against infection

Effective measures must be taken to protect against infection in line with laboratory guidelines.

Instructions for disposal

The package must be disposed of in accordance with the current disposal guidelines.

Used solutions and solutions that are past their shelf-life must be disposed of as special waste in accordance with local guidelines. Information on disposal can be obtained under the Quick Link "Hints for Disposal of Microscopy Products" at www.microscopy-products.com. Within the EU the currently applicable REGULATION (EC) No 1272/2008 on classification, labelling and packaging of substances and mixtures, amending and repealing. Directives 67/548/EEC and 1999/45/EC, and amending Regulation (EC) No 1907/2006 applies.

Auxiliar	y reagei	nts	
Cat. No.	100206	Phenol GR for analysis ACS, Reag. Ph Eur	250 g, 1 kg
Cat. No.	100327	Hydrochloric acid in ethanol for microscopy	1 I, 5 I
Cat. No.	100497	Tb-color modified Staining kit for the detection of myco- bacteria (AFB) by hot staining method	1 set
Cat. No.	100579	DPX new non-aqueous mounting medium for microscopy	500 ml
Cat. No.	100971	Ethanol 96% suitable for use as excipient EMPROVE® exp Ph Eur,BP	1 l, 2.5 l, 10 l
Cat. No.	100974	Ethanol denatured with about 1 % methyl ethyl ketone for analysis EMSURE®	1 I, 2.5 I
Cat. No.	101287	Löffler's methylene blue solution for microscopy	100 ml, 500 ml, 2.5 l
Cat. No.	102560	ISOSLIDE® AFB Control Slides with reference tissue for the detection of Mycobacteria in histological tissue	25 tests
Cat. No.	104699	Immersion oil for microscopy	100-ml dropping bottle, 100ml, 500 ml
Cat. No.	107961	Entellan® new rapid mounting medium for microscopy	100 ml, 500 ml, 1 l
Cat. No.	108000	Sputofluol® for microbiology and microscopy	11
Cat. No.	108298	Xylene (isomeric mixture) for histology	4
Cat. No.	108562	Aquatex® (aqueous mounting agent) for microscopy	50-ml dropping bottle
Cat. No.	109016	Neo-Mount® anhydrous mounting medium for microscopy	100-ml dropping bottle, 500 ml
Cat. No.	109843	Neo-Clear® (xylene substitute) for microscopy	51
Cat. No.	115943	Methylene blue (C.I. 52015) for microscopy Certistain®	25 g, 100 g
Cat. No.	116450	Tb-color staining kit for the microscopic investigation of mycobacteria (cold staining)	1 set

Hazard classification

Cat. No. 1.05226.0100

Please observe the hazard classification printed on the label and the information given in the safety data sheet.

The safety data sheet is available on the website and on request.

Main components of the product

Cat. No. 1.05226.0100 C.I. 42520

 $\mathsf{C}_{22}\mathsf{H}_{24}\mathsf{CIN}_3$ M = 365.91 g/mol

Other IVD products

Cat. No. 101597 TB-Fluor phenol-free 1 set Staining kit for the acid-free smear

examination with fluorescence microscopy

(Auramin-Rhodamine staining)

Cat. No. 109204 Giemsa's azur eosin methylene blue 100 ml, 500 ml,

solution 1 I, 2.5 I

for microscopy

Histosec® pastilles Cat. No. 111609 1 kg, 10 kg (4x soldification point 56-58°C 2.5 kg), 25 kg

embedding agent for histology

Cat. No. 111885 Gram-color 1 set

stain set for the Gram staining method

Literature

- 1. Romeis Mikroskopische Technik, Editors: Mulisch, Maria, Welsch, Ulrich, 2015, Springer-Verlag Berlin Heidelberg
- 2. Theory and Practice of Histological Techniques, John D Bancroft and Marilyn Gamble, 6th Edition
- Theory and application of Microbiological Assay, Hewitt, W. and Vincent, S., 1989, Academic Press
- 4. Conn's Biological Stains: A Handbook of Dyes, Stains and Fluorochromes for Use in Biology and Medicine, 10th Edition, (ed. Horobin, R.W. and Kiernan, J.A). Bios, 2002







Catalog number



Consult instructions for use

Manufacturer

accompanying documents

Use by YYYY-MM-DD



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Caution, consult

Merck KGaA, 64271 Darmstadt, Germany Tel. +49(0)6151 72-2440 www.microscopy-products.com

