Sustainable Alternatives to Triton™ X-100 Detergent for Biomanufacturing: The Deviron® Detergent Portfolio



Alice Antonello, Sarah Lechat, Anja Licht, Almut Rapp, Kakolie Banerjee, Sandra Johnson, Corinne Miller, Henry George, Agnieszka Lass-Napiorkowska, Angeles Mecate-Zambrano, Alexa Prager, Michelle Zöller, Vanessa Lotis, Thibaut Deschamps, Sandy McNorton

Process Solutions, Merck Life Science KGaA Process Solutions, MilliporeSigma

Introduction

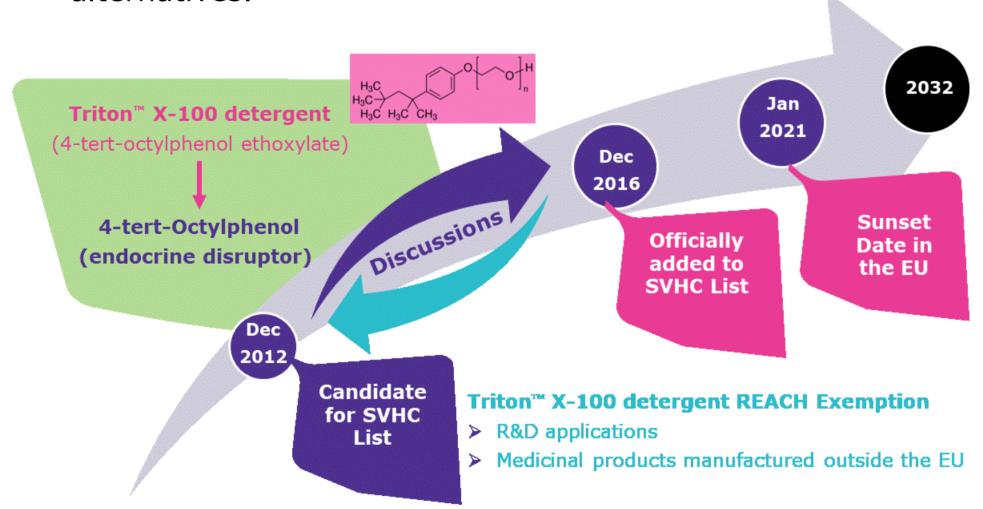
Viral safety is a **major concern for biotherapeutic** manufacturers.

- <u>Cell-based processes</u> may produce endogenous retroviral particles, and adventitious viruses can be introduced from contaminated source materials or during the manufacturing process.
- <u>Human plasma-derived products</u> are at risk of containing viruses, despite extensive screening of donation material.

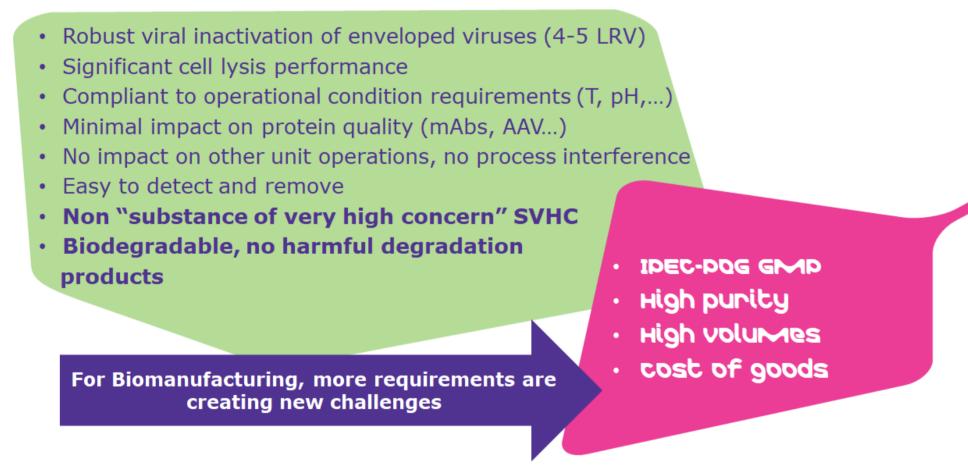
Detergent-mediated **viral inactivation** is widely used in multiple biotherapeutic production processes as part of an overall virus safety strategy.

Triton™ X-100 detergent is widely used.

- A degradation product of Triton™ X-100 detergent is 4tert-octylphenol, an endocrine disruptor hazardous for the environment.
- Triton™ X-100 detergent was classified as "Substance of Very High Concern SVHC" in the REACH (Registration, Evaluation, Authorization and Restriction of Chemicals) in 2017.
- The European Chemical Agency (ECHA) prohibited the unauthorized use of Triton™ X-100 detergent in the EU in January 2021.
- The biotherapeutic industry together is faced with the challenges of identifying, producing and implementing new alternatives.



Minimum criteria for alternative detergent candidates

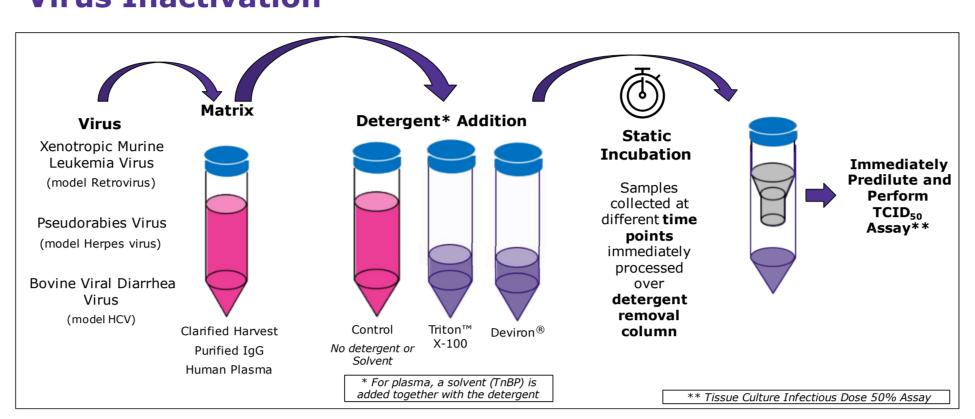


We offer a **portfolio of detergents**, **the Deviron**[®] detergents, to meet different application and process requirements.

Deviron® Detergents

	Deviron® C-16 detergent	Deviron® 13-S9 detergent	
Chemical name	N,N-Dimethyltetradecylamin- N-oxide	Alcohols, C11-15-secondary, ethoxylated	
CAS number	3332-27-2	68131-40-8	
Surfactant	Zwitterionic (pI 8.9)	Non-ionic	
СМС	0.002-0.003 wt % (24 °C)	0.005 wt % (24 °C)	
Form	30 % wt. water solution	Pure substance (100 % wt.)	
Biodegradability (OECD 301B)	Readily biodegradable	Readily biodegradable	
Toxicology report	Available	Available	
Quality marker	ISO9001	IPEC-PQG GMP	
Documentation package	Emprove® Evolve MQ400	Emprove® Expert MQ500	
Viral inactivation efficiency	Yes >5 LRV	Yes >5 LRV	
Cell lysis efficiency	Yes	Yes	
Endotoxin removal for plasmid purification	Yes	Yes	
Detection method	HPLC-ELSD method available	HPLC-ELSD method available	

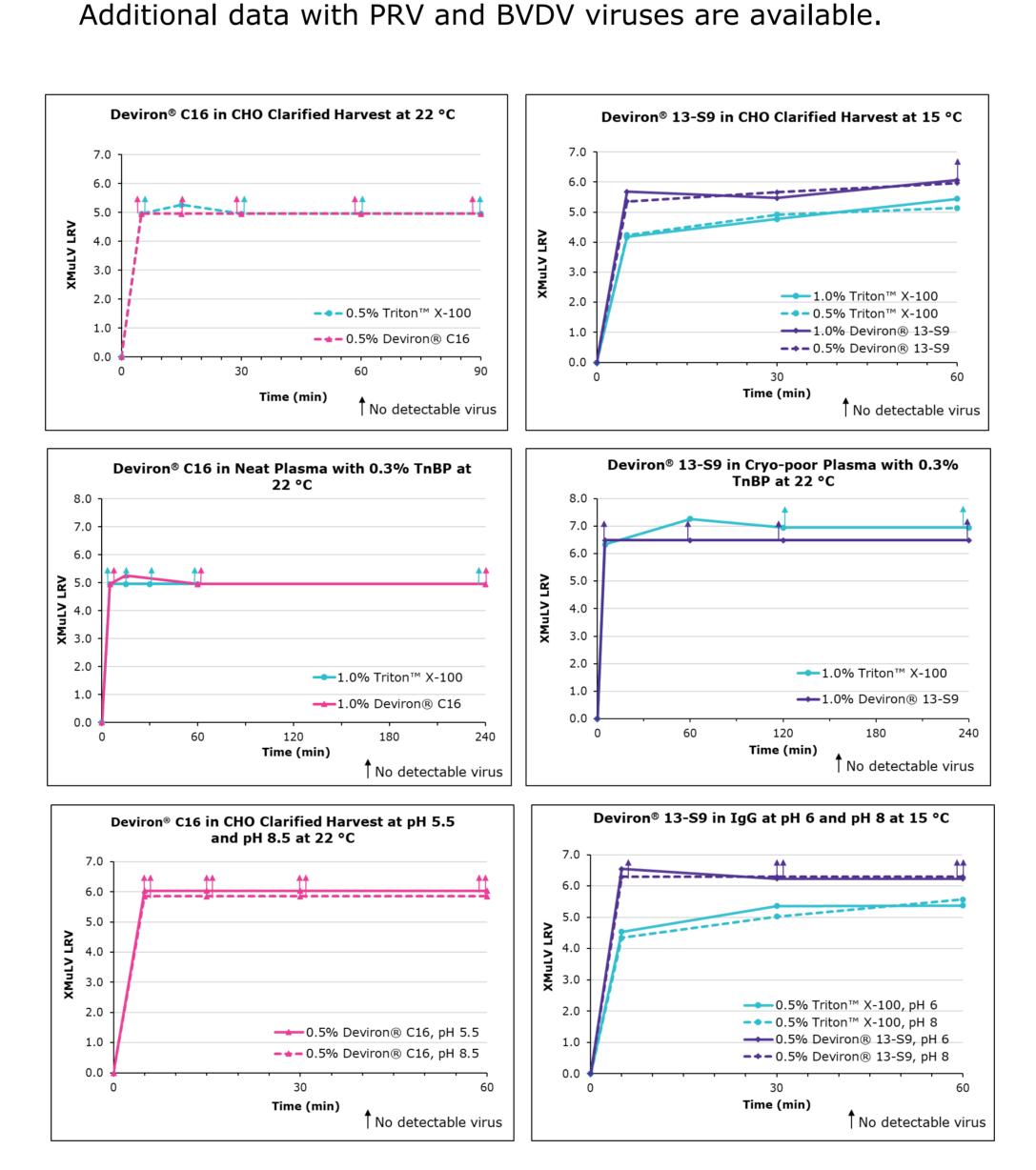
Virus Inactivation



Virus inactivation was assessed in mAb-containing CHO clarified harvest and human plasma matrices with **XMuLV** model.

The standard viral inactivation practice for **mAb processes** is described in the **ASTM E3042-16**. In this procedure the **detergent** concentration is ≥ 0.5 %, no solvent (i.e., TnBP) and incubation time ≥ 60 min.

Plasma processes typically utilize 1.0 % detergent and 0.3 % solvent TnBP and longer incubation time (4-6 h). Deviron® C16 and Deviron® 13-S9 detergents demonstrate effective viral inactivation (LRV > 5) in all conditions tested.



Cell Lysis

HEK293 and Sf-RVN® cells were cultivated according to their type, suspension or adherent, in the appropriate cell culture media.

HEK293 cells were transfected after one day of culture with the selected plasmid polyethylenimine (PEI) complexation and Sf-RVN® cells infected with Baculovirus at the time of seeding. After cultivation, the cells were lysed.

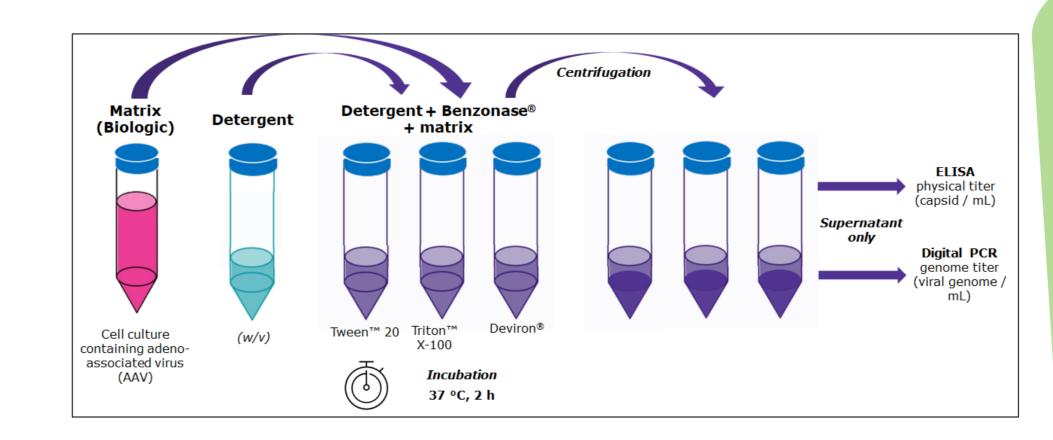
Detergent: 0.5 % wt.

Nuclease: 25 U/mL Benzonase® endonuclease with 2 mM MgCl₂

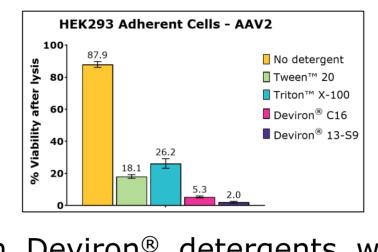
Lysis time: 2 h

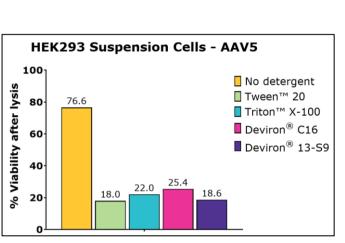
Temperature: 37 °C

Following incubation, total cell count and viability values determined. Virus-containing supernatant was clarified by centrifugation. The supernatant was analyzed to determine physical titer and genome titer. After detergent removal, the AAV infectivity was measured.

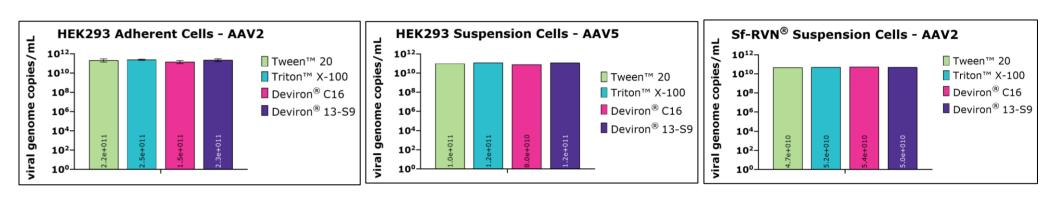


Deviron® Detergents

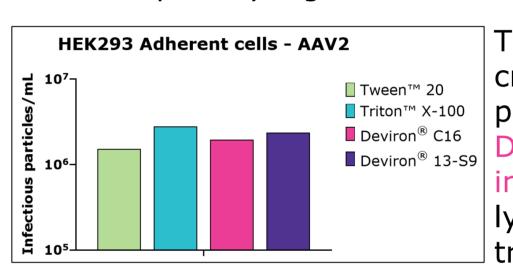




Both Deviron® detergents were excellent in lysing HEK293 and Sf-RVN® cells (data not shown, microscopic images show complete cell lysis). Both Deviron® detergents were comparable or even better than the benchmarks.



Both Deviron® detergents were as efficient as the benchmarks in releasing capsids and the viral genome titers were comparably high.



The infectivity of the AAV is a crucial parameter for final products. Both of the Deviron® detergents preserve infectivity of the AAV after the lysis, as evaluated with the transduction unit assay.

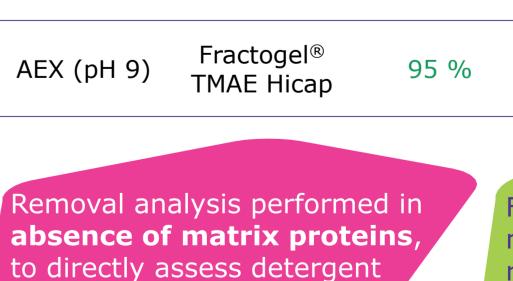
The Deviron® detergents are an advantageous alternative to the benchmarks, combining efficacy, sustainability, and easier handling.

Removal of Deviron® detergents in Downstream Process
Effective detergent removal from a process stream needs to
be achieved by downstream steps. Removal was assessed
with several different chromatographic resins.

Deviron® C16 detergent

Columns tested	Type of resin	Remova
Protein A	Eshmuno [®] A	99.6 %
CEX (pH 6)	Eshmuno® S	43-64 %
AEX (bind/elute mode pH 9)	Fractogel® TMAE Hicap	99.8 %
AEX (flow through mode pH 9)	Fractogel® TMAE Hicap	Not suitable
	c point (pI) Dev detergent = 8. 9	
•	.9 → net positiv 9 → net negativ	
Deviron [©]	® 13-S9 dete	rgent
Columns tested	Type of resin	Removal
Protein A	Eshmuno® A	100 %

99 %

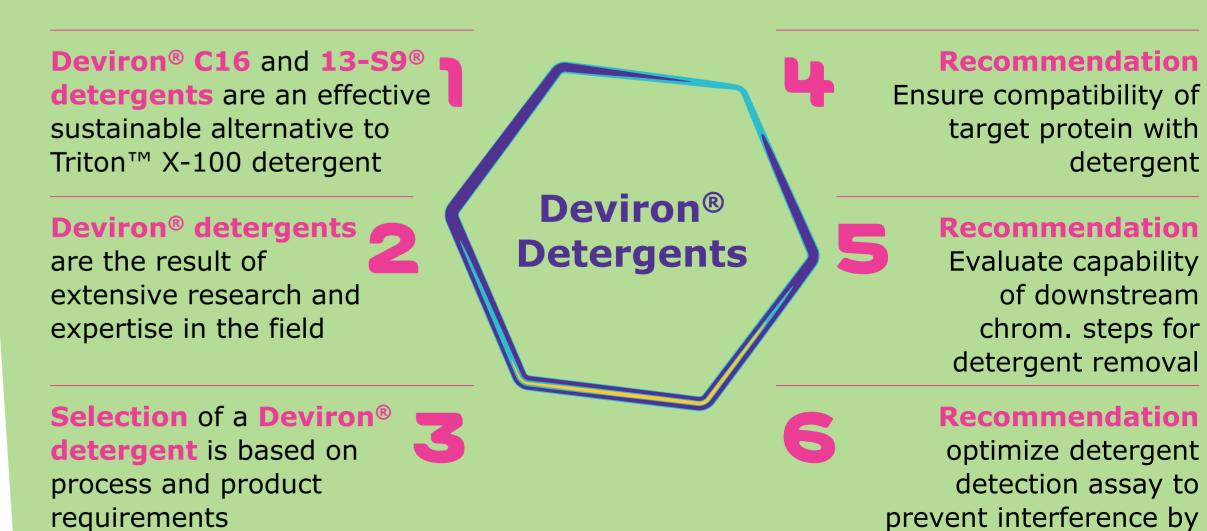


CEX (pH 6) Eshmuno® S

binding to resin

For specific process and method development, reach out for technical support

matrix proteins



The Life Science business of Merck operates as MilliporeSigma in the U.S. and Canada.