

# Quantitative Analysis of Urea in Milk by Thin-Layer Chromatography using TLC Explorer

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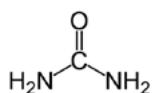
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## Abstract

Milk adulteration presents a significant health hazard to humans. One of the most frequently encountered adulterants in milk is urea as it mimics a higher protein content. However, such adulteration presents a significant health hazard to humans. Here a simple and efficient approach by TLC is demonstrated providing a rapid and accurate detection of urea in milk using a new TLC analysis and documentation system, the TLC-Explorer.

## Introduction

Milk is one of the most crucial general food and nutrition products, as it is rich in calcium and other essential nutrients, including proteins, fats, minerals, carbohydrates, and various vitamins needed by both infants and adults.<sup>1,2</sup> Being an affordable source of high-protein nutrition, government nutrition support programs in many countries provide milk and dairy products free of charge to students in schools, addressing nutritional deficiencies and by that benefiting future generations.<sup>3</sup> This has led to increased demand for milk over the years. To maximize profits and exploit the gap between demand and supply, some vendors engage in unethical practices, such as adulterating milk to artificially increase the value of their provided product.<sup>4</sup> Adulteration with nitrogen rich compounds such as urea has become an easy and cheap practice of such vendors to imitate a higher protein content and increase solid-not-fat (SNF) content.<sup>5</sup> However, the these adulterants poses significant health risks for consumers as they can adversely affect various organs in the human body, including the heart, liver, kidneys, and intestines, and may even result in death.<sup>6,7</sup>



Urea, also known as carbamide, is an organic compound that plays a crucial role in the metabolism of nitrogen-containing compounds in animals and is the primary nitrogenous substance found in the urine of mammals. The urea content in cow's milk typically ranges from 20 mg to 70 mg per 100 mL, and levels exceeding this range are considered adulterated or intentionally added.<sup>8</sup>

In this work a simple and efficient analytical method for determination and quantification of urea in milk using TLC is demonstrated for two commercial milk samples. The samples are diluted, centrifuged and then directly applied onto a silica gel 60 F<sub>254</sub> thin layer chromatography (TLC) plate. The identification, quantification, and imaging of urea present in the samples is performed with a TLC Explorer instrument (Figure 1).

This system is an easy-to-use hardware and software concept for TLC plate imaging, analysis, and data handling. It is applicable for routine measurements, such as in-process control, method development, or daily research activities.



Figure 1. TLC Explorer (left -closed; right – plate tray open).

The TLC Explorer system contains a camera and three different LED light sources to view, analyze and document TLC plates at wavelengths of 245 nm, 366 nm, and VIS (white light). It is a “no service maintenance required” instrument, that has a check window for a quick and safe visual examination of chromatograms/plates and the safety feature of no exposure of UV light outside the device during measurements. The instrument is equipped with a Wi-Fi antenna making it easy to connect with any desktop device (read more at [SigmaAldrich.com/tlc-explorer](https://SigmaAldrich.com/tlc-explorer)).

The performance of the here developed method on the TLC Explorer is also partially assessed regarding response as per the ICH Q2 R2 guidelines.<sup>9</sup>

## Experimental

### Reagent preparation

**Ehrlich reagent:** Dissolve 5 g of 4-(dimethylamino) benzaldehyde in 40 mL ethanol and slowly add 10 mL HCl (37%) with cooling. The temperature should not fall below 20 °C or rise above 40 °C.

### Standard Preparation

- **Diluent:** Methyl alcohol and water (4:1 v/v)
- **Urea standard solution (0.7 mg/mL):** Weigh and transfer 7 mg of urea certified reference material to 10 mL volumetric flask, add 5 mL of diluent and sonicate for 5 min with intermittent shaking, then make up to volume with diluent, and mix thoroughly. Use different application volumes for calibration curve creation.

### Sample Preparation

Two milk samples (1 & 2) were sourced from a local supermarket.

- **Milk sample solution:** Dissolve 5 mL of milk in 5 mL of water, mix thoroughly and centrifuged and at 2500 rpm for 10 min. Use the clear supernatant and spot on the chromatographic plate (5 µL).
- **Recovery sample solution:** Weigh and transfer 7 mg of urea certified reference material to 10 mL volumetric flask, add 5 mL of milk sample 1, sonicate for 5 min with intermittent shaking, then make up to volume with diluent, and mix thoroughly, and centrifuged and at 2500 rpm for 10 minutes. The clear supernatant was used at different application volumes.

**Note:** The milk samples used for spiking did not show detectable background of urea.

### TLC Method

The separation was performed on silica gel 60 F<sub>254</sub> on aluminum support (**Table 1**). After drying the plate was sprayed with Ehrlich reagent and then viewed white light.

**Table 1.** HPTLC conditions

HPTLC Parameter	
Chromatographic conditions	
TLC plate:	Silica gel 60 F <sub>254</sub> 20 x 20 cm, TLC plate (1.05554)
Plate pretreatment:	Prewash the plate with methanol and allow to dry in air before use
Sample application:	See <b>Table 2</b>
Sample application (auto spotter):	Camag Linomat 5 using band length = 4 mm, track distance = 13 mm, distance from left edge = 15 mm, distance from lower edge = 20 mm. Allow to dry completely before plate development. Track listing in <b>Table 2</b>
Mobile phase:	1-Propanol:water 9:1 (v:v)
Chamber conditions:	Unsaturated
Migration distance:	15 cm (3/4 of plate)
Drying:	100 °C
Staining/ derivatization:	Spray with Ehrlich reagent (4-(dimethylamino) benzaldehyde 5 g/50 mL), dry in a current of cold air, and immediately examine under white light
Detection:	White light and 366 nm

**Table 2.** Applied spots and application volume for quantification (linearity and recovery)

Track No.	Spot Name used in TLC Explorer	Application volume (µL)
1	Urea calibration standard - 1.4 µg/spot	2
2	Urea calibration standard - 2.1 µg/spot	3
3	Urea calibration standard - 2.8 µg/spot	4
4	Urea calibration standard - 3.5 µg/spot	5
5	Urea calibration standard - 4.2 µg/spot	6
6	Urea calibration standard - 4.9 µg/spot	7
7	Milk sample 1 - spot 1	5
8	Milk sample 1 - spot 2	5
9	Milk sample 2 - spot 1	5
10	Milk sample 2 - spot 2	5
11	Recovery sample solution with 1.4 µg/spot	2
12	Recovery sample solution with 2.8 µg/spot	4
13	Recovery sample solution with 4.2 µg/spot	6
14	Urea calibration standard - 2.8 µg/spot	4

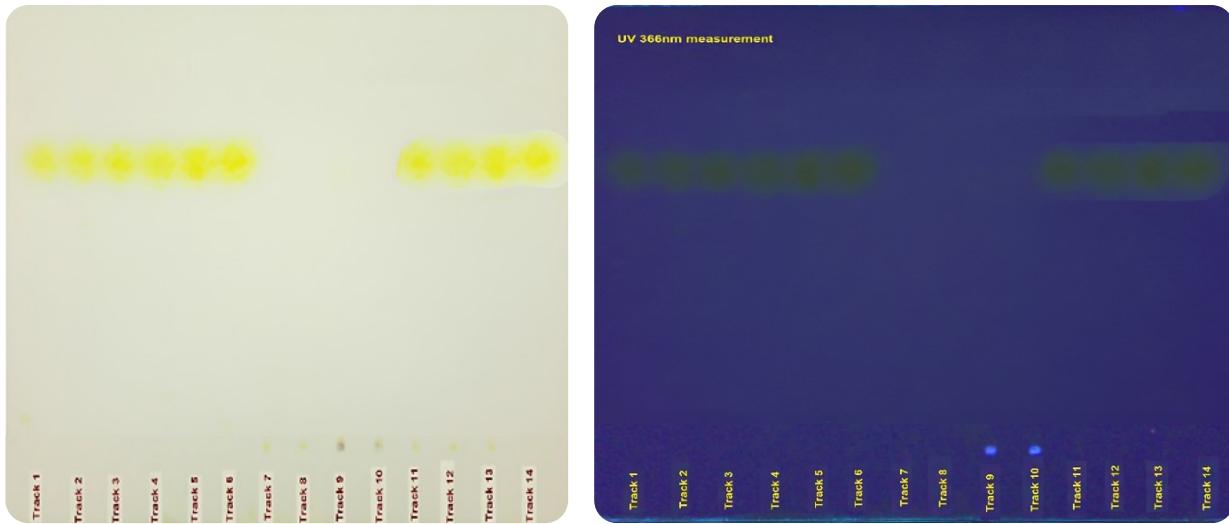
## Results and Discussion

Calibration standards and milk samples were analysed by evaluation of the developed intense yellow urea bands after derivatization and video densitometry under white light (**Figure 2**). An example video densitogram, created by the TLC-Explorer system, for the calibration track with 4.9 µg urea per spot is shown in **Figure 3**.

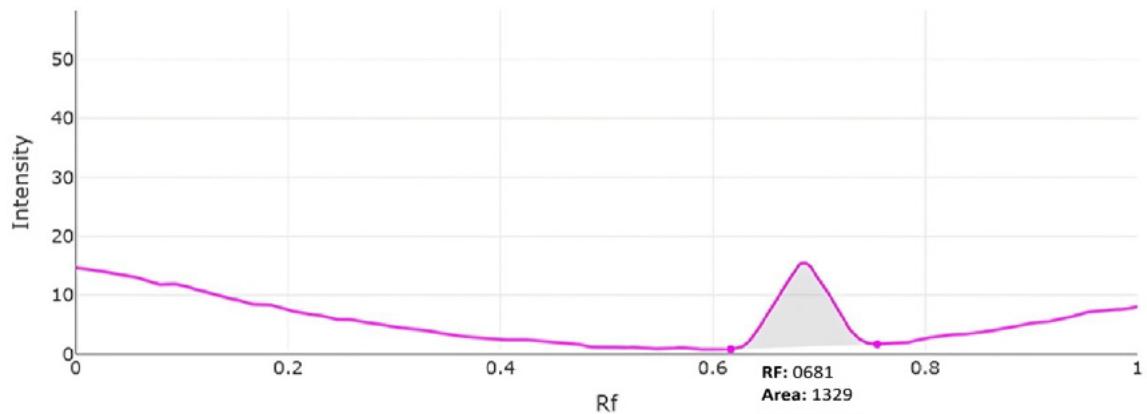
The experimental results such as retention factors ( $R_f$ ) and peak areas determined for the applied spots are shown in **Table 3**.

The calibration ranged from 1.4 µg per spot to 4.9 µg per spot (6 calibrants, tracks 1-6), representing for a 5 µL application of diluted milk sample, concentrations in the original milk of 560 to 1960 µg/mL (56 to 196 mg/100 mL). The polynomial calibration curve fit provided an  $R^2$  value of 0.9964 (**Figure 4**).

Using three different volumes of the spiked recovery sample showed recoveries in the range of 87.1 to 103.9% (**Table 3**).



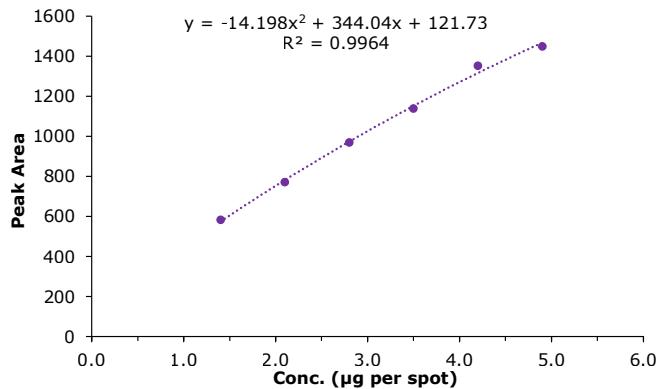
**Figure 2.** TLC chromatograms for identification of urea in milk samples (left: visible light; right: UV 366 nm). Tracks 1 to 6 were utilized for the determination of linearity in the concentration range of urea ranging from 1.4 µg per spot to 4.9 µg per spot, tracks 7 and 8 represent commercial milk sample 1 and tracks 9 and 10 display the results for commercial milk sample 2. Tracks 12 to 14 were used for recovery studies with individually spiking of 1.4, 2.8 and 4.2 µg per spot individually over the milk sample spots. Track 14 shows the result of the analysis of urea urea standard solution spot (2.8 µg). (Tables 2 & 3)



**Figure 3.** Representative video densitogram for the 4.9 µg per spot applied standard solution (track 6).

**Table 3.** Results for calibration, quantification, retention factors, and recovery of urea (see also Figures 2, 3 and 4)

Experimental parameter	Track no.	Concentration (µg/spot)	R <sub>f</sub>	Peak area
Calibration Standards	1	1.4	0.689	567
	2	2.1	0.684	752
	3	2.8	0.680	952
	4	3.5	0.684	1,059
	5	4.2	0.677	1,258
	6	4.9	0.681	1,329
Milk sample 1	7	Not detected	Not detected	Not detected
	8	Not detected	Not detected	Not detected
Milk sample 2	9	Not detected	Not detected	Not detected
	10	Not detected	Not detected	Not detected
Recovery study	11	Recovery sample solution with	0.679	558.8 (Recovery 87.1%)
	12	1.4 µg/spot (2 µL sample)	0.681	932.8 (Recovery 103.9%)
	13	Recovery sample solution with	0.675	1,167.8 (Recovery 94.5.1%)
Urea standard solution	14	2.8 µg/spot (4 µL sample)	0.681	948



**Figure 4.** Calibration curve (polynomial) for urea in µg per spot vs peak area in the concentration range of 1.4 µg to 4.9 µg per spot (see also Table 3).

## Conclusion

A method for the determination of urea in milk by TLC with derivatization and video densitometry using the TLC- Explorer analysis and documentation system was presented.

This developed method showed sufficient retention ( $R_f$  0.68) and was evaluated in the concentration range from 1.4 µg per spot to 4.9 µg per spot, representing milk sample concentrations of 56 to 196 mg/100 mL. The calibration showed an  $R^2$  value of 0.9964 and spiking experiments showed recoveries ranging from 87.1 to 103.9%.

Thus, the here displayed method showed that TLC plates along with the TLC Explorer, are a viable option for a quick and efficient limit check for urea content in milk samples.

## References

1. Paula C. Milk nutritional composition and its role in human health, Nutrition, Volume 30, Issue, 2014 June 6; (30):619-627 ,DOI: <https://doi.org/10.1016/j.nut.2013.10.011>
2. Shelver W, Lupton, S, Shappell N, Smith J, Hakk H. Distribution of Chemical Residues among Fat, Skim, Curd, Whey, and Protein Fractions in Fortified, Pasteurized Milk. ACS Omega 2018; 3: 8697–8708. DOI: <https://doi.org/10.1021/acsomega.8b00762>
3. Griffin M. Overview of Worldwide School Milk Programmes, 3rd International School Milk Conference, FAO, Kunming, 2005; pp 1–10. [https://www.fao.org/fileadmin/templates/est/COMM\\_MARKETS\\_MONITORING/Dairy/Documents/Overview\\_of\\_worldwide\\_school\\_milk\\_programmes.pdf](https://www.fao.org/fileadmin/templates/est/COMM_MARKETS_MONITORING/Dairy/Documents/Overview_of_worldwide_school_milk_programmes.pdf)
4. Harsha B, Suman K. Gap in Supply and Demand: Impact on Milk Safety. International Journal of Agriculture and Biological Sciences, 2021October 31, DOI:10.5281/zenodo.5734154
5. Shalileh F, Sabahi H, Dadmehr M, Hosseini M. Sensing approaches toward detection of urea adulteration in milk. Microchemical journal, devoted to the application of micro techniques in all branches of science. 2023;193(108990):108990. <http://dx.doi.org/10.1016/j.microc.2023.108990>.
6. Das S, Goswami B, Biswas K. Milk adulteration and detection: A review. Sensor letters. 2016;14(1):4–18. <http://dx.doi.org/10.1166/sl.2016.3580>
7. Reddy M, Venkatesh K, Venkata C, Reddy S. Adulteration of Milk and Its Detection: A Review. Int. J. Chem. Stud. 2017; 5: 613–617. <https://www.chemijournal.com/archives/2017/vol5issue4/PartJ/5-3-158-426.pdf>
8. van Duinkerken G et al. Milk urea concentration as an indicator of ammonia emission from dairy cow barn under restricted grazing. Journal of dairy science. 2011;94(1):321–335. <http://dx.doi.org/10.3168/jds.2009-2263>.
9. ICH Harmonised Guideline ICH Q2 (R2) – Validation of Analytical Procedures. [https://database.ich.org/sites/default/files/ICH\\_Q2%28R2%29\\_Guideline\\_2023\\_1130.pdf](https://database.ich.org/sites/default/files/ICH_Q2%28R2%29_Guideline_2023_1130.pdf)

## Related Products

Description	Cat. No.
<b>HPTLC Plate, TLC Instrument</b>	
TLC aluminum sheets, Silica gel 60 F <sub>254</sub> , 20 cm × 20 cm, aluminum support, Pk.25	1.05554
TLC Explorer, digital TLC analysis and documentation device	1.52610
<b>Solvents, Reagents, and Standards</b>	
Ultrapure water from Milli-Q® ultrapure water purification system	ZIQ7005T0C
Ethyl alcohol (Pure 190 proof, ACS spectrophotometric grade, 95.0%)	493511
1-Propanol for liquid chromatography LicChrosolv®	1.01024
4-(Dimethylamino)benzaldehyde GR for analysis Reag. Ph Eur	1.03058
Urea, Pharmaceutical Secondary Standard; Certified Reference Material	PHR1406

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